

#### Prioritizing Variants in Personal Genomes: Using functional impact & recurrence, with particular application to cancer

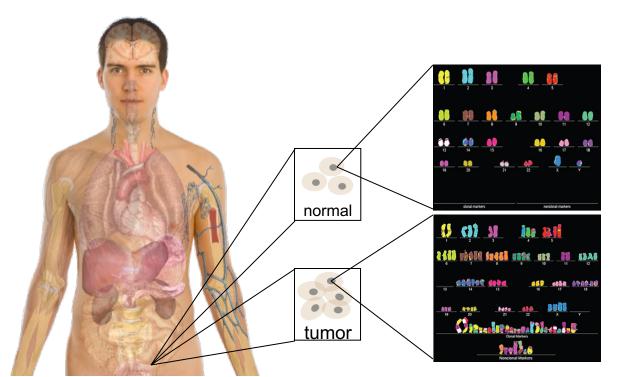
Mark Gerstein Yale

Slides freely downloadable from Lectures.GersteinLab.org & "tweetable" (via @MarkGerstein). No Conflicts for this Talk

See last slide for more info.

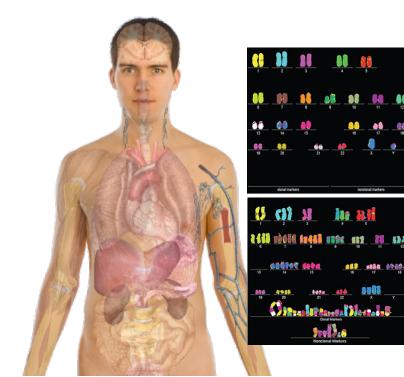
## Personal Genomics as a Gateway into Biology

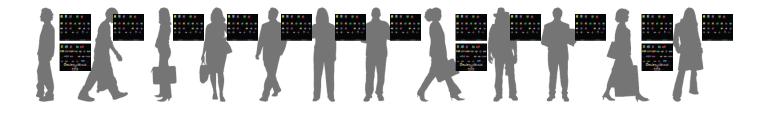
Personal genomes will soon become a commonplace part of medical research & eventually treatment (esp. for cancer). They will provide a primary connection for biological science to the general public.

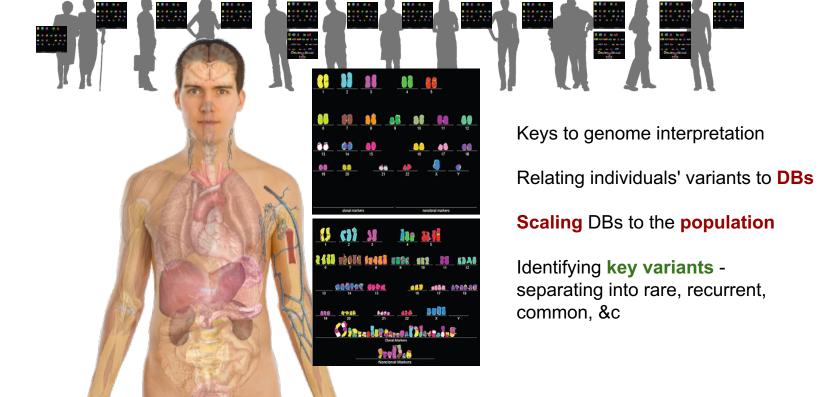


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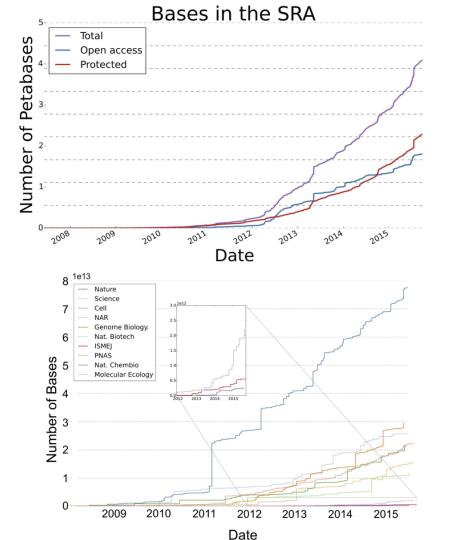






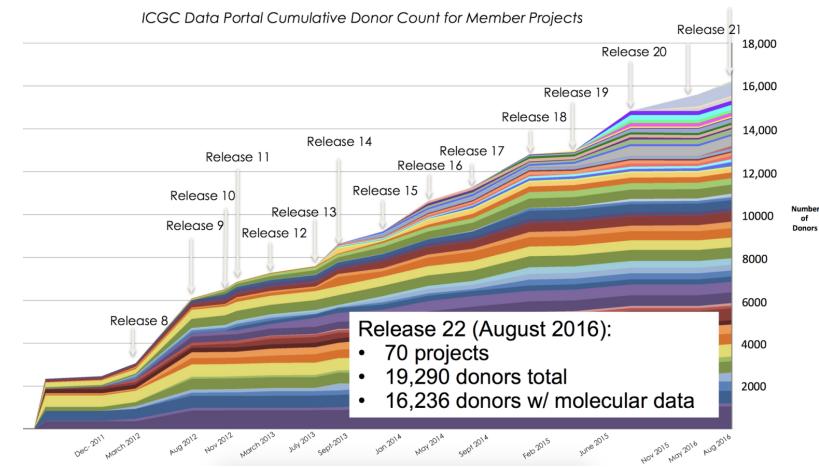
#### DB Growth: explosion of data scale & a diversity of uses

- The type of sequence data deposited has changed as well.
  - Protected data represents an increasing fraction of all submitted sequences.



## **Growth of ICGC datasets**



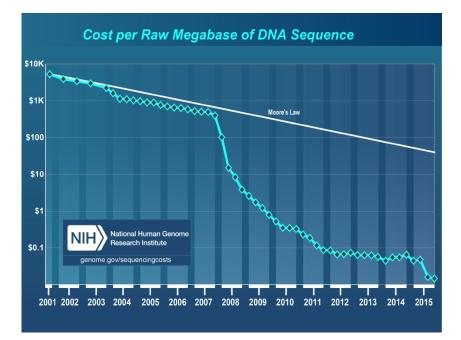


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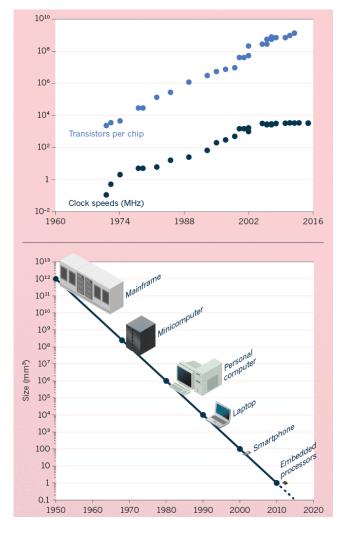
#### Sequencing Data Explosion: Faster than Moore's Law?

- In the early 2000's, improvements in Sanger sequencing produced a scaling pattern similar to Moore's law.
- The advent of NGS was a shift to a new technology with dramatic decrease in cost).



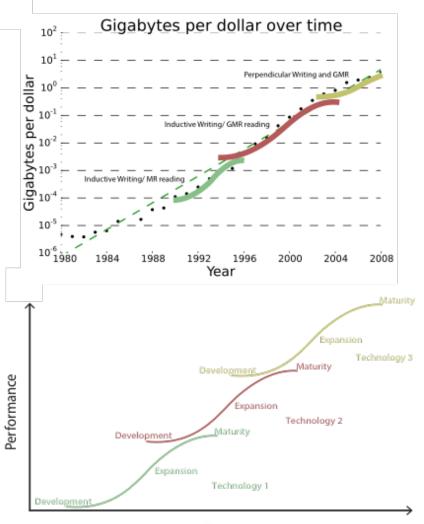
#### Moore's Law: Exponential Scaling of Computer Technology

- Exponential increase in the number of transistors per chip.
- Led to improvements in speed and miniaturization.
- Drove widespread adoption and novel applications of computer technology.

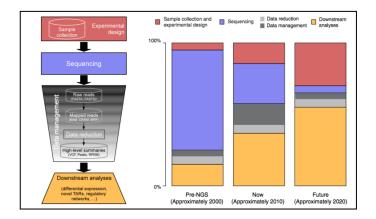


#### Kryder's Law and S-curves underlying exponential growth

- Moore's & Kryder's Laws
  - As important as the increase in computer speed has been, the ability to store large amounts of information on computers is even more crucial
- Exponential increase seen in Kryder's law is a superposition of S-curves for different technologies

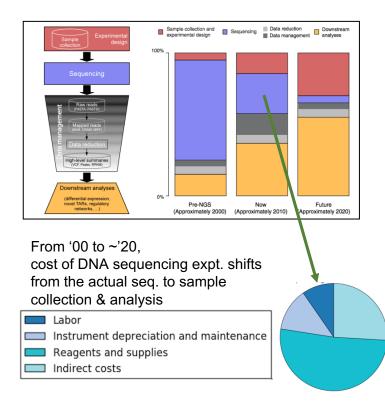


Time

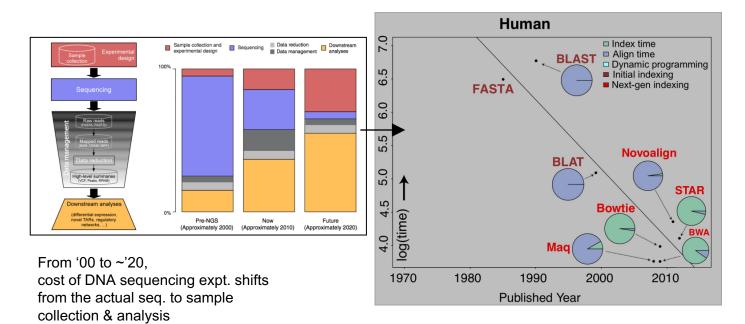


From '00 to ~'20, cost of DNA sequencing expt. shifts from the actual seq. to sample collection & analysis

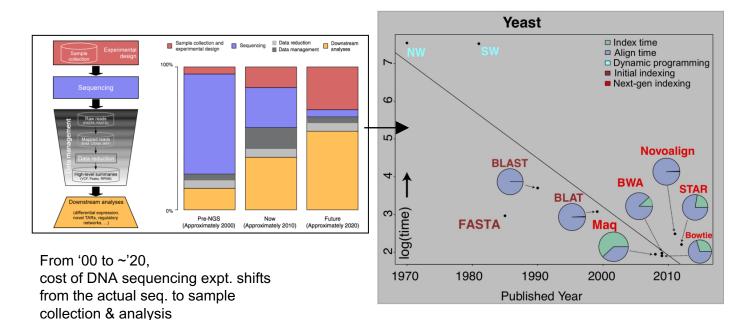
[Sboner et al. ('11), Muir et al. ('15) Genome Biology]



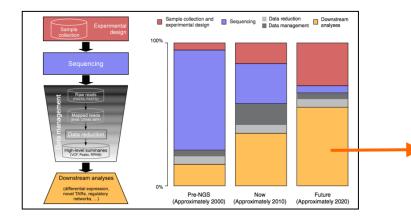
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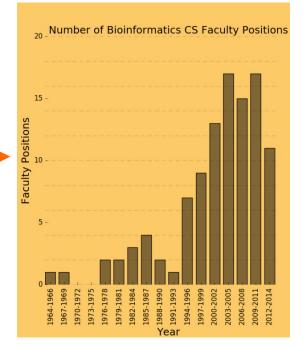
Alignment algorithms scaling to keep pace with data generation



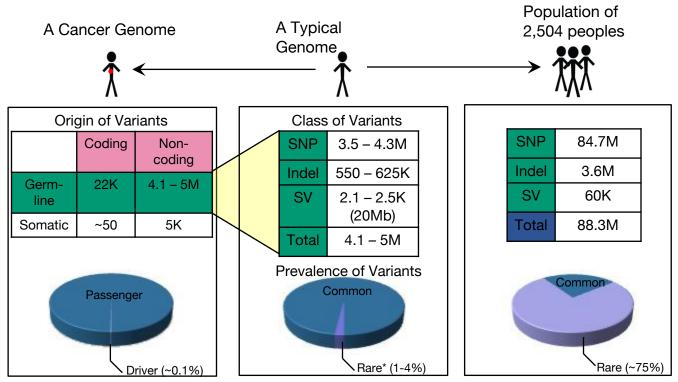
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## **Human Genetic Variation**



\* Variants with allele frequency < 0.5% are considered as rare variants in 1000 genomes project.

The 1000 Genomes Project Consortium, Nature. 2015. 526:68-74 Khurana E. et al. Nat. Rev. Genet. 2016. 17:93-108

## Finding Key Variants

### Germline



#### Common variants

- · Can be most readily associated with phenotype (ie disease) via GWAS
- Usually their functional effect is weaker
- Many are non-coding
- Issue of LD in identifying the actual causal variant.

#### Rare variants

- Associations are usually underpowered due to low frequencies but often have larger functional impact
- Can be collapsed in the same element to gain statistical power (burden tests).

## Finding Key Variants

**Somatic** 



#### Overall

• Often these can be thought of as very rare variants

#### Drivers

- Driver mutation is a mutation that directly or indirectly confers a selective growth advantage to the cell in which it occurs.
- A typical tumor contains 2-8 drivers; the remaining mutations are passengers.

#### Passengers

• Conceptually, a passenger mutation has no direct or indirect effect on the selective growth advantage of the cell in which it occurred.

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  - LoF annotation as a complex problem + finding deleterious LoFs
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  - <u>FunSeq</u> integrates evidence, with a "surprisal" based weighting scheme
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  - Recurrence #2:

#### (Low-power) application to **pRCC**

- WGS finds additional facts on the canonical driver, MET. Other suggestive non-coding hotspots.
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## Variant Annotation Tool (VAT), developed for 1000G FIG

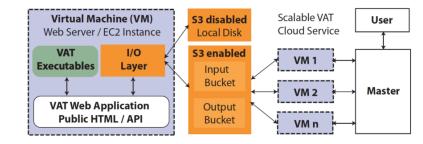
#### VCF Input

Output:

- Annotated VCFs
- Graphical representations of functional impact on transcripts

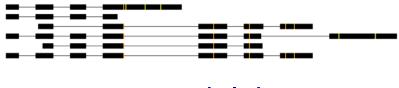
#### Access:

- Webserver
- AWS cloud instance
- Source freely available



**CLOUD APPLICATION** 





vat.gersteinlab.org

Habegger L.\*, Balasubramanian S.\*, et al. Bioinformatics, 2012

### **Complexities in LOF annotation**

Transcript isoforms, Isoform 1 distance to stop, Case 1 Isoform 2 functional domains, Affects only Isoform 1 protein folding, Isoform 1 etc. Reference Isoform 2 Affects both isoforms Balasubramanian S. et al., Genes Dev., '11 Balasubramanian S.\*, Fu Y.\* et al., NComms., '17 Isoform 1 Case 2 Isoform 2 SLC2A2 1KG ENST00000469787 ENST00000497642 HGMD ENST0000382808 ENST0000314251

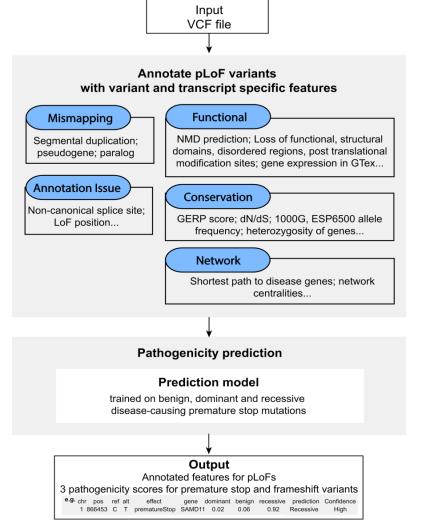
Impact of a SNP on alternate splice forms

## <u>Annotation of</u> <u>Loss-of-Function</u> <u>Transcripts</u> (ALoFT)

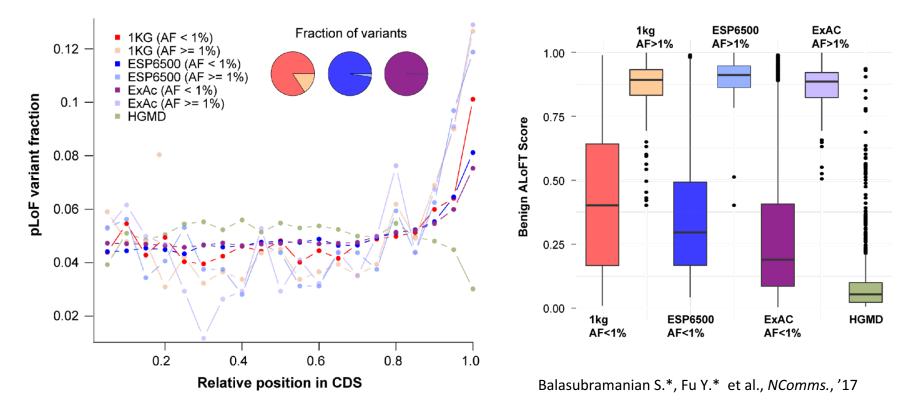
Runs on top of VAT

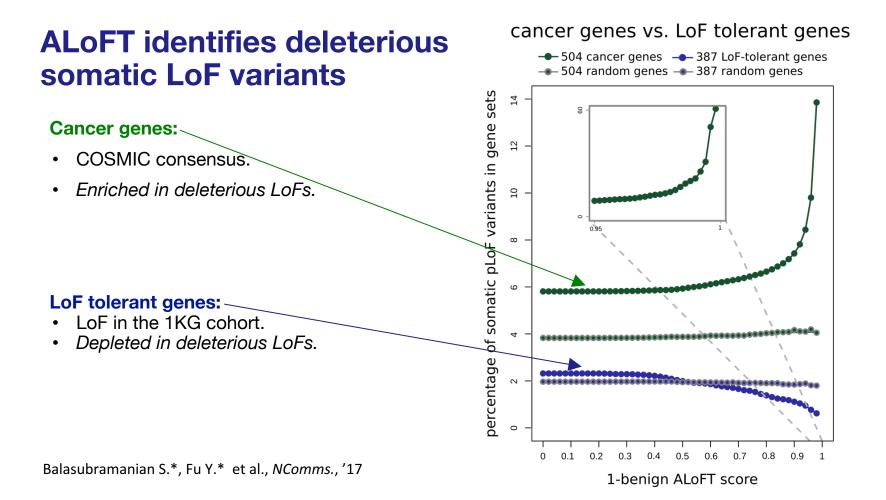
Output:

- Impact score: benign or deleterious.
- Decorated VCF.

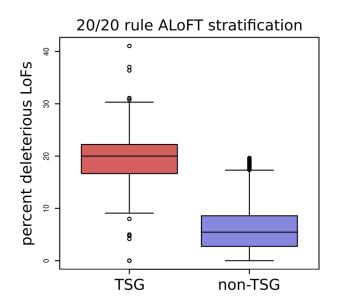


## LoF distribution varies as expected by mutation set (from healthy people v from disease)





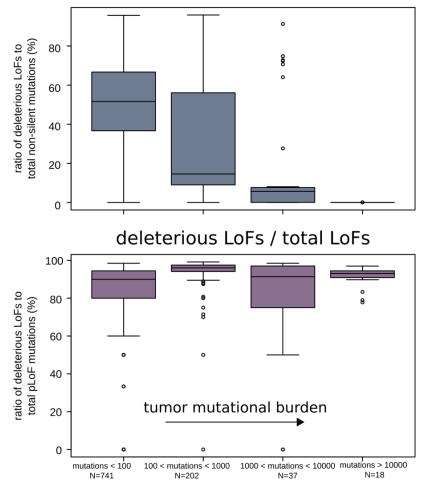
#### **ALoFT refines cancer** mutation characterization



*Vogelstein et al.* '13: if >20% of mutations in gene inactivating  $\rightarrow$  tumor suppressor gene (TSG).

ALoFT further refines 20/20 rule predictions.

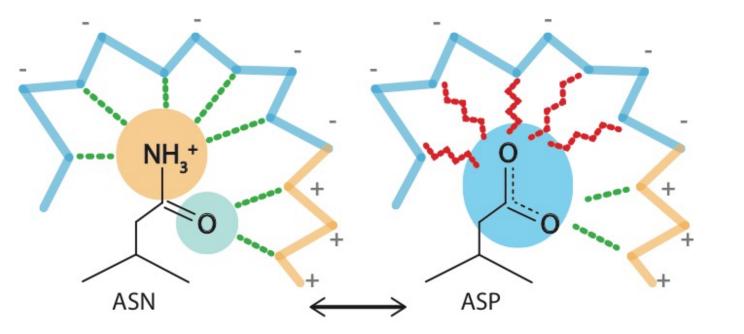
deleterious LoFs / total mutations



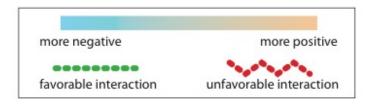
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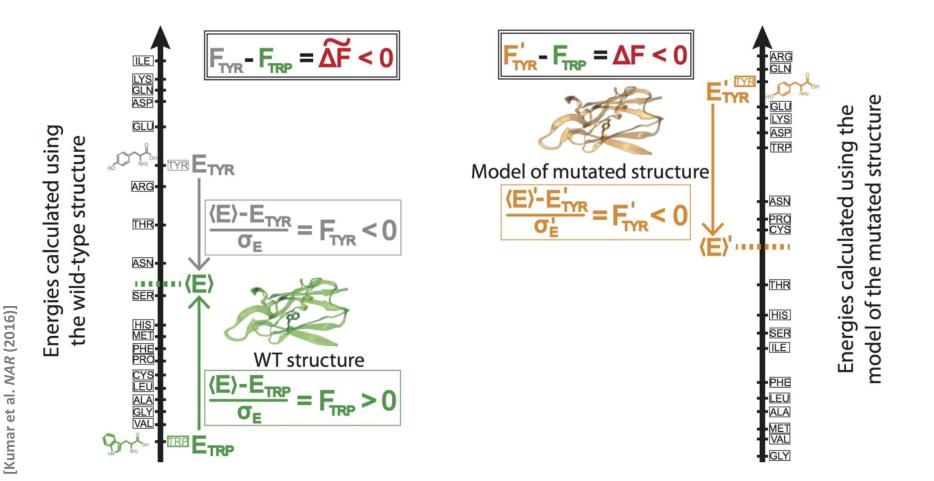
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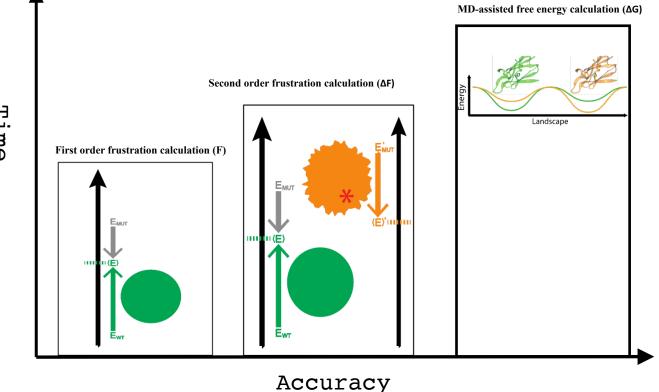
### What is localized frustration ?



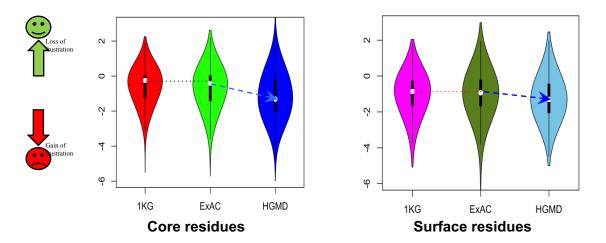
## Workflow for evaluating localized frustration changes (ΔF)



## **Complexity of the second order frustration calculation**

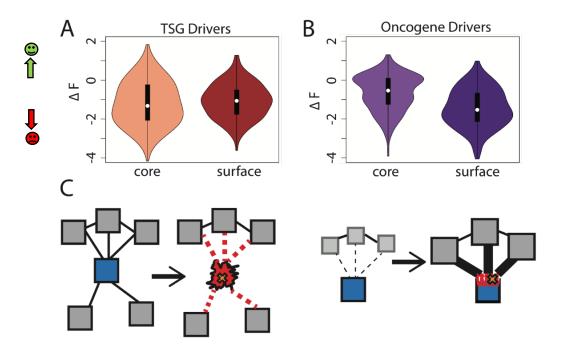


## Comparing ΔF values across different SNV categories: disease v normal



Normal mutations (1000G) tend to unfavorably frustrate (less frustrated) surface more than core, but for disease mutations (HGMD) no trend & greater changes

## Comparison between ΔF distributions: TSGs v. oncogenes



[Kumar et al, *NAR* (2016)]

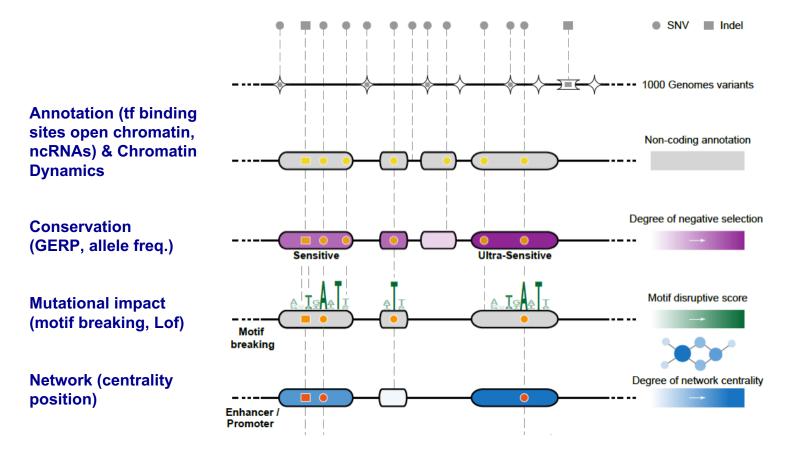
SNVs in TSGs change frustration more in core than the surface, whereas those associated with oncogenes manifest the opposite pattern. This is consistent with differences in LOF v GOF mechanisms.

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# Funseq: a flexible framework to determine functional impact & use this to prioritize variants

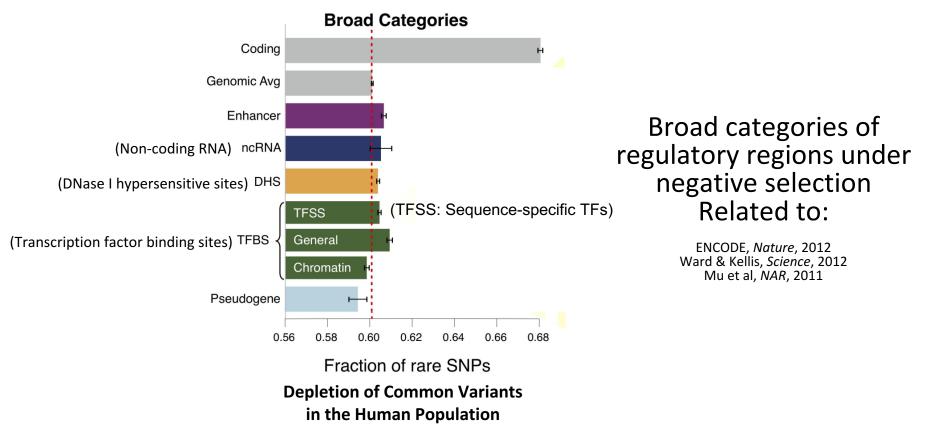


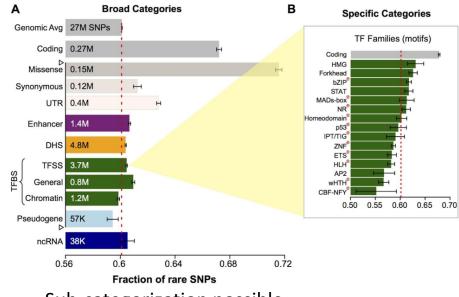
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#### Finding "Conserved" Sites in the Human Population:

#### Negative selection in non-coding elements based on Production ENCODE & 1000G Phase 1

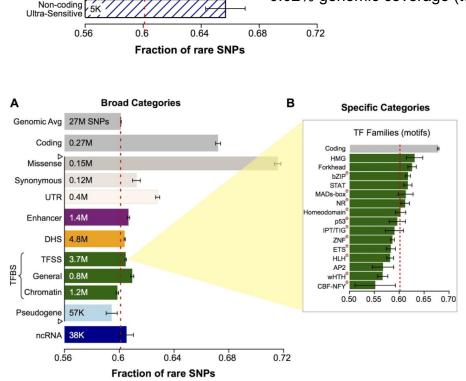




Differential selective constraints among specific sub-categories

Sub-categorization possible because of better statistics from 1000G phase 1 v pilot

[Khurana et al., Science ('13)]



~0.4% genomic coverage (~ top 25)

~0.02% genomic coverage (top 5)

Sub-categorization possible because of better statistics from 1000G phase 1 v pilot

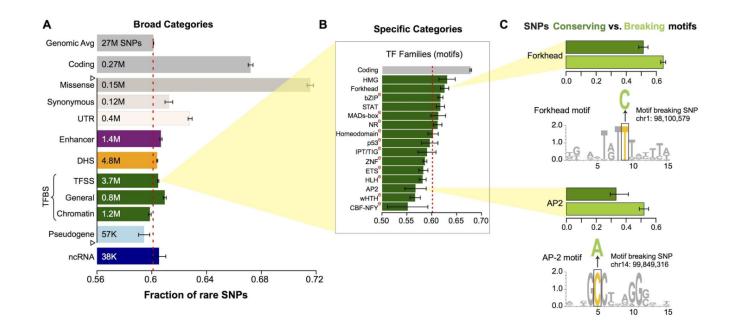
Non-coding

Defining Sensitive non-coding Regions

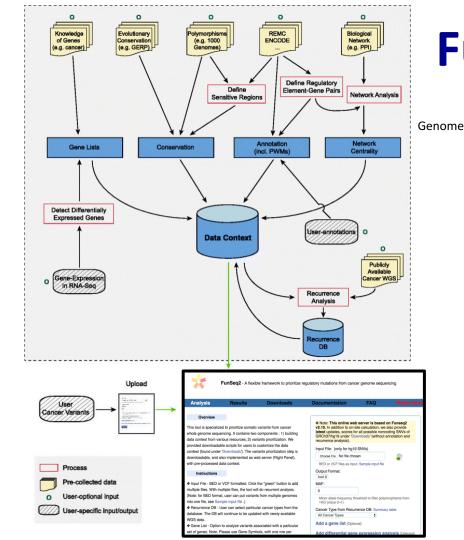
Start 677 highresolution noncoding categories; Rank & find those under strongest selection

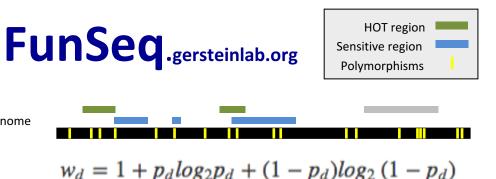
[Khurana et al., Science ('13)]

#### SNPs which break TF motifs are under stronger selection



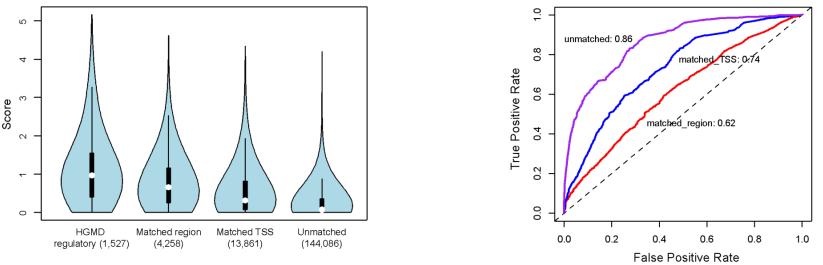
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- Info. theory based method (ie annotation "surprisal") for weighting consistently many genomic features
- Practical web server
- Submission of variants & precomputed large data context from uniformly processing large-scale datasets

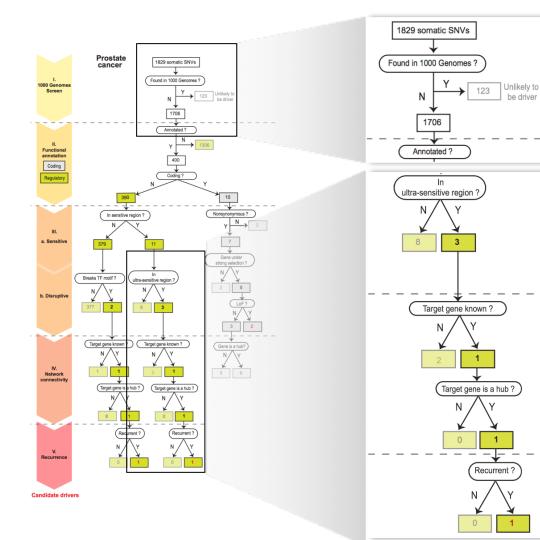
# Germline pathogenic variants show higher core scores than controls



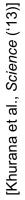
- 3 controls with natural polymorphisms (allele frequency >= 1%)
  - 1. Matched region: 1kb around HGMD variants
  - 2. Matched TSS: matched for distance to TSS
  - 3. Unmatched: randomly selected

Ritchie et al., Nature Methods, 2014

[Fu et al., GenomeBiology ('14, in revision)]



Flowchart for 1 Prostate Cancer Genome (from Berger et al. '11)



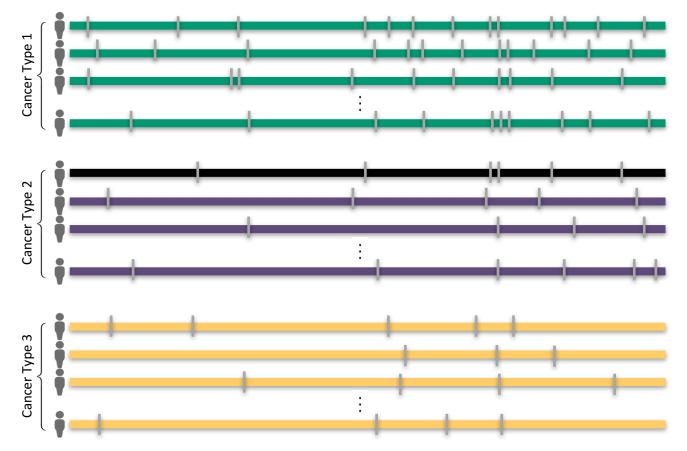
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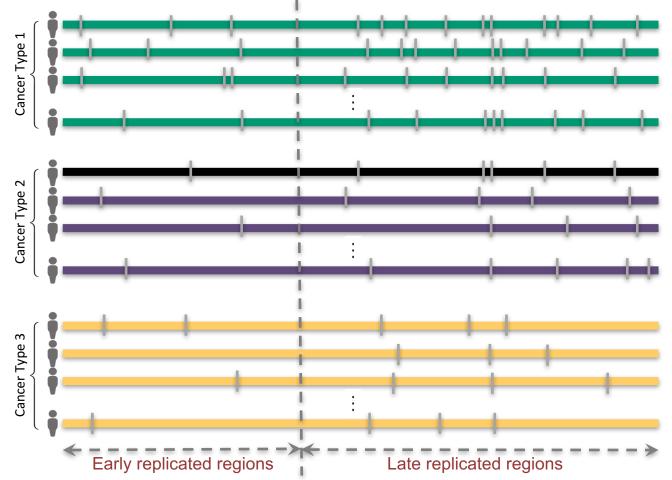
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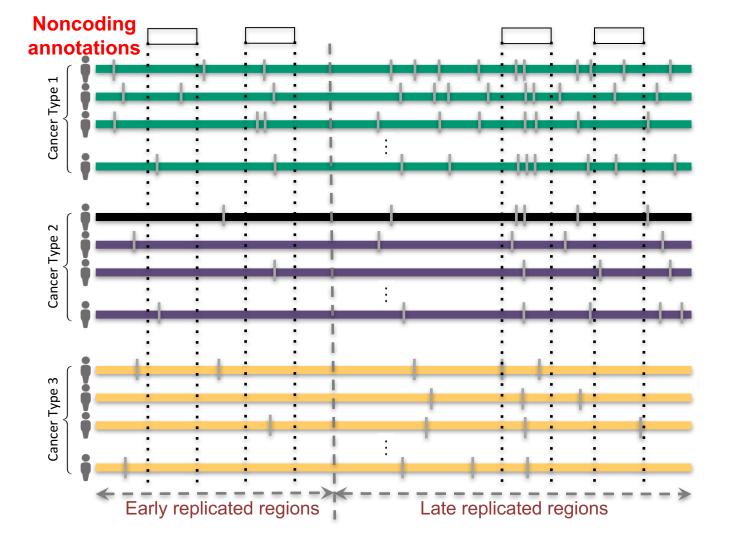
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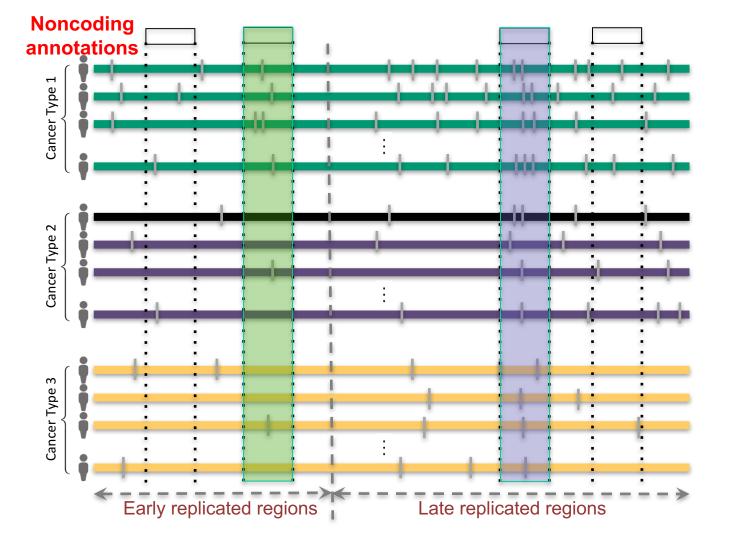
### **Mutation recurrence**

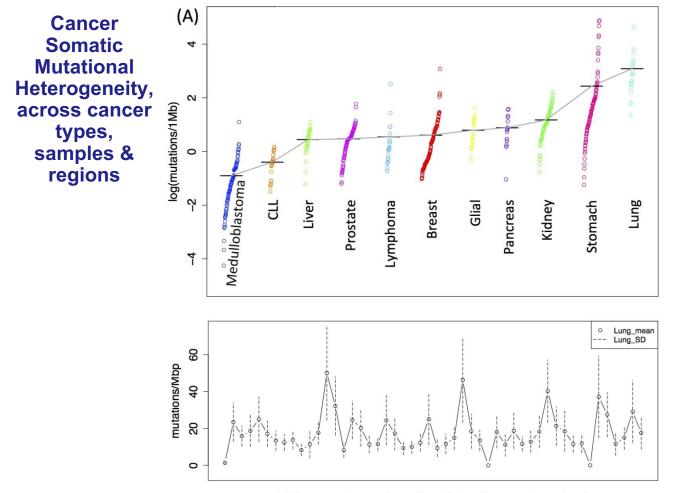


## **Mutation recurrence**

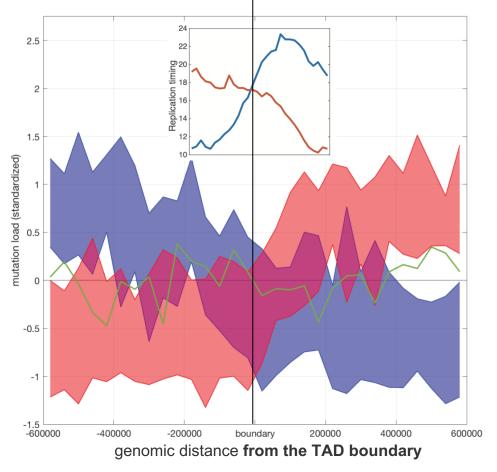


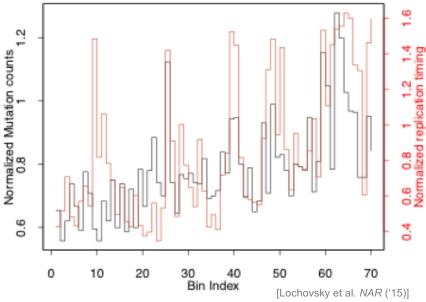






1 Mbp genome regions (locations chosen at random)



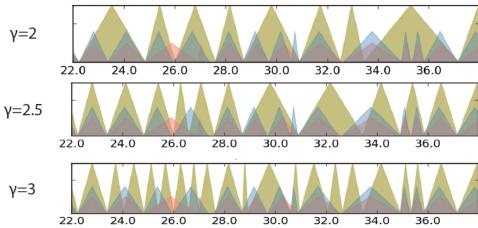


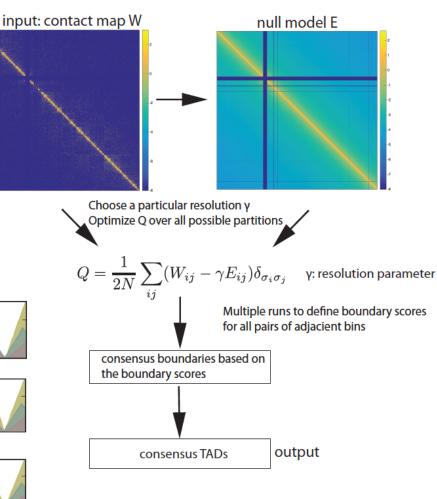
Chromatin remodeling failure leads to more mutations in early-replicating regions

Variation in somatic mutations is closely associated with chromatin structure (TADs) & replication timing

### mrTADFinder:

#### Identifying TADs at multiple resolutions by maximizing modularity vs appropriate null





[Yan et al., PLOS Comp. Bio. ('17)]

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### **Cancer Somatic Mutation Modeling**

#### PARAMETRIC MODELS

Model 1: Constant Background Mutation Rate (Model from Previous Work)  $x_i : Binomial(n_i, p)$ 

# Model 2a: Varying Mutation Rate with Single Covariate Correction

- $x_i$ : Binomial $(n_i, p_i)$
- $p_i$ : Beta $(\mu | R_i, \sigma | R_i)$
- $\mu | R_i, \sigma | R_i$  : constant within the same covariate rank

#### Model 2b: Varying Mutation Rate with Multiple Covariate Correction $x_i$ : Binomial $(n_i, p_i)$

- $p_i$ : Beta $(\mu | \mathbf{R}_i, \sigma | \mathbf{R}_i)$
- $\mu | \mathbf{R}_i, \sigma | \mathbf{R}_i$ : constant within the same covariate rank

- Suppose there are k genome elements. For element i, define:
  - n;: total number of nucleotides
  - x; the number of mutations within the element
  - -p: the mutation rate
  - $-R_i$ : the covariate rank of the element
  - Non-parametric model is useful when covariate data is missing for the studied annotations
    - Also sidesteps issue of properly identifying and modeling every relevant covariate (possibly hundreds)

#### NON-PARAMETRIC MODELS

Assume constant background mutation rate in local regions.

#### Model 3a: Random Permutation of Input Annotations

Shuffle annotations within local region to assess background mutation rate.

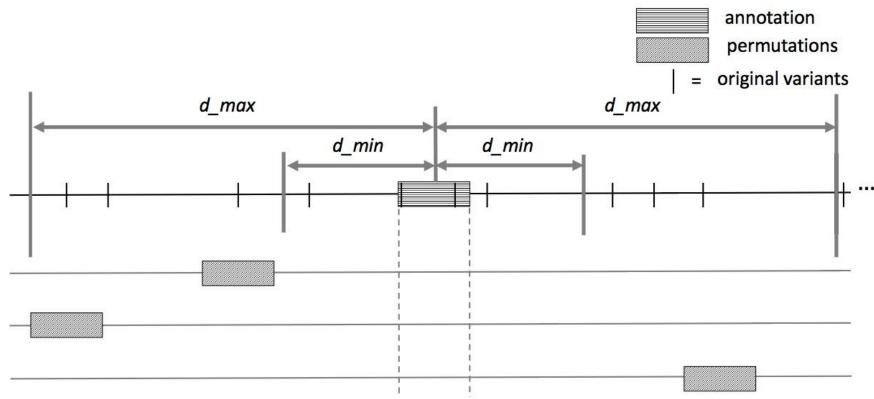
#### Model 3b: Random Permutation of Input Variants

Shuffle variants within local region to assess background mutation rate.

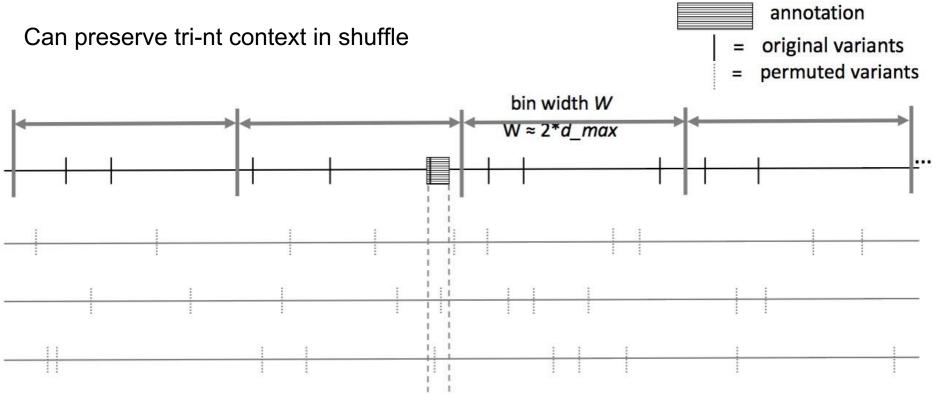
[Lochovsky et al. Bioinformatics in press]

[Lochovsky et al. NAR ('15)]

## **MOAT-a: Annotation-based permutation**



# **MOAT-v: Variant-based Permutation**



[Lochovsky et al. Bioinformatics in press]

# **MOAT-s: a variant on MOAT-v**

11

43

• A somatic variant simulator

Shuffling variants

5

Given a set of input variants, shuffle to new locations, taking genome structure into account

original variantspermuted variants

...

...

...

...

Binning whole genome Marking equivalence classes (bins with similar covariate vectors) Overlaying variants (with tri-nucleotide indexing) 1 2 2 333 3 3 4444444 55 6 7

344 43

.....

4 2

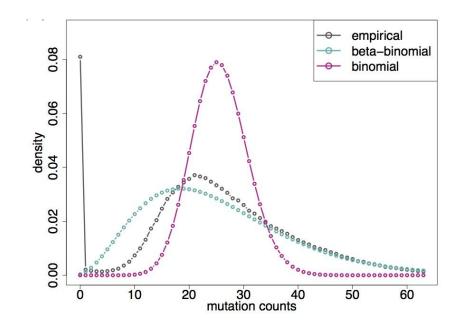
6

5

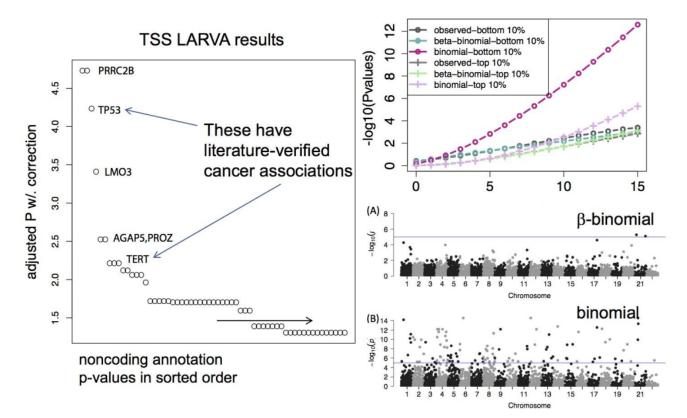
[Lochovsky et al. *Bioinformatics* in press]

# **LARVA Model Comparison**

- Comparison of mutation count frequency implied by the binomial model (model 1) and the beta-binomial model (model 2) relative to the empirical distribution
- The beta-binomial distribution is significantly better, especially for accurately modeling the over-dispersion of the empirical distribution



## **LARVA Results**



# MOAT: recapitulates LARVA with GPU-driven runtime scalability

Gene Name	Documented role with cancer	Pubmed ID	
SLC3A1	Cysteine transporter SLC3A1 promotes breast cancer tumorigenesis	28382174	
ADRA2B	reduce cancer cell proliferation, invasion, and migration 2502635		
SIL1	subtype-specific proteins in breast cancer	23386393	
TCF24	NA	NA	
AGAP5	significant mutation hotspots in cancer 252		
TMPRSS13	Type II transmembrane serine proteases in cancer and viral infections	19581128	
ERO1L	Overexpression of ERO1L is Associated with Poor Prognosis of Gastric Cancer	26987398	

MOAT's high mutation burden elements recapitulate LARVA's results & published noncoding cancer-associated elements.

Computational efficiency of MOAT's NVIDIA<sup>™</sup> CUDA<sup>™</sup> version, with respect to the number of permutations, is dramatically enhanced compared to CPU version.

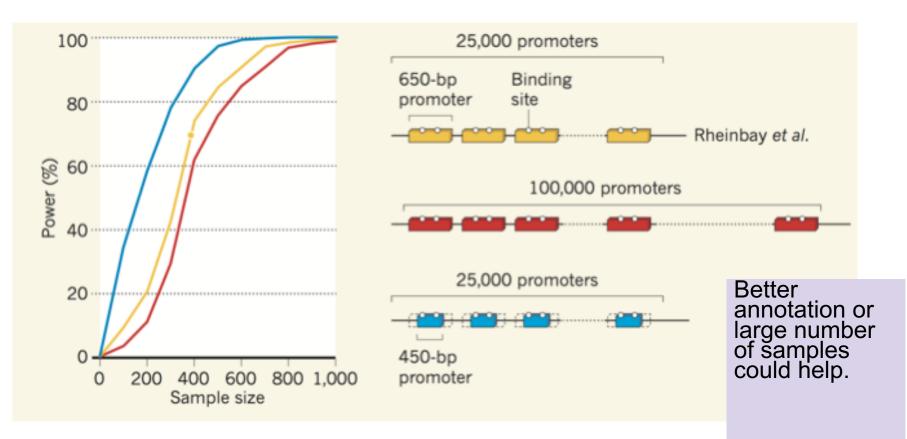
Number of permutations	Fold speedup of CUDA version
1k	14x
10k	100x
100k	256x

Prioritizing Variants in Personal Genomes: Using functional impact & recurrence, with particular application to cancer

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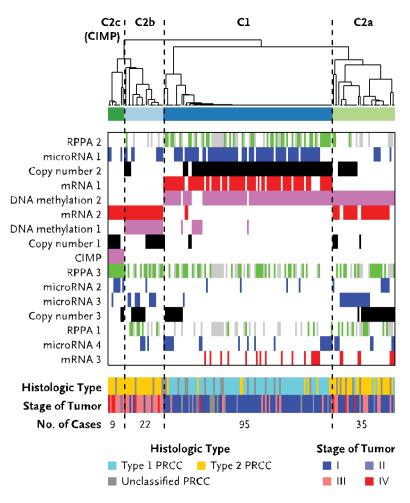
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  - Background mutation rate significantly varies & is correlated with replication timing & TADs
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  - (Low-power) application to **pRCC** 
    - WGS finds additional facts on the canonical driver, MET. Other suggestive non-coding hotspots.
    - Analysis of signatures & tumor evolution helps identify key mutations in different ways

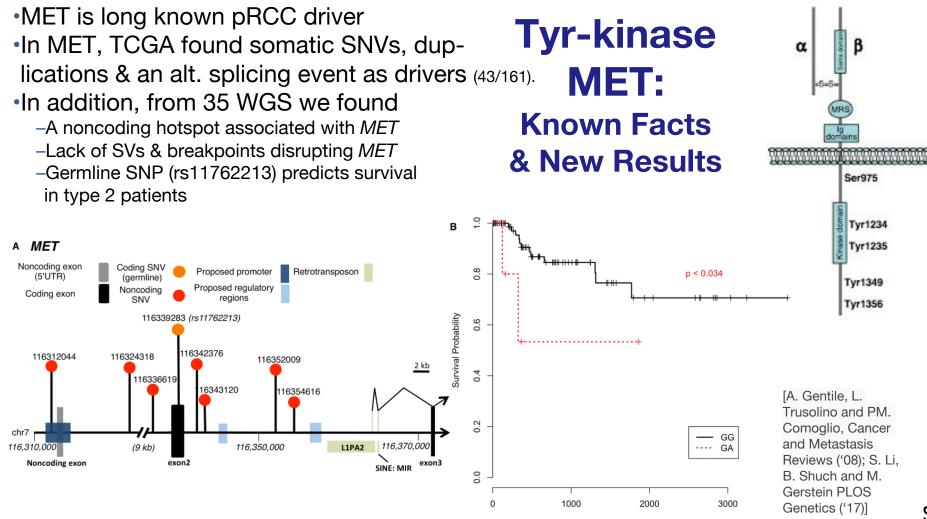
# Power, as an issue in driver discovery



# An (underpowered) case study: pRCC

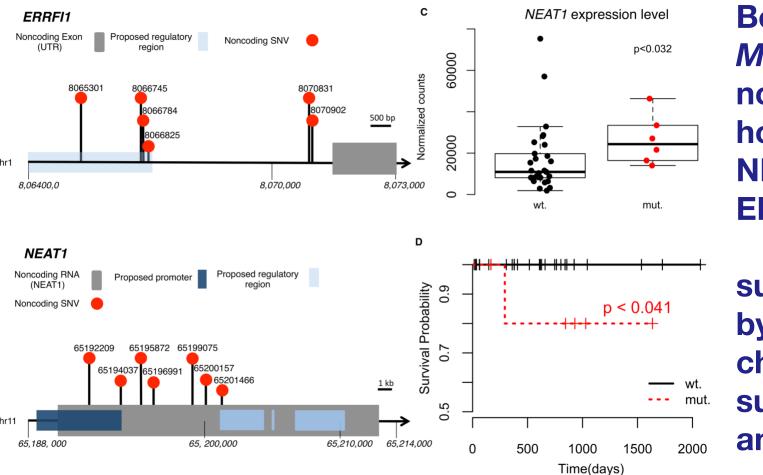
- Kidney cancer lifetime risk of 1.6% & the papillary type (pRCC) counts for ~10% of all cases
- TCGA project sequenced 161 pRCC exomes & classified them into subtypes
  - Yet, cannot pin down the cause for a significant portion of cases....
  - •35 WGS of TN pairs, perhaps useful? But not that definitive from a recurrence perspective





Time(days)

60 = Lectures.GersteinLab.org

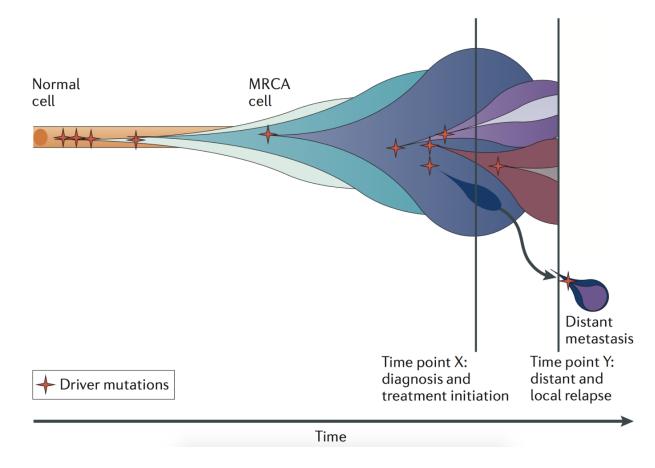


### Beyond *MET*: 2 non-coding hotspots in NEAT & ERRFI1,

supported by expr. changes & survival analysis

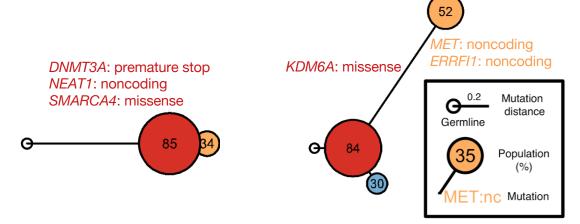
61 = Lectures.GersteinLab.org

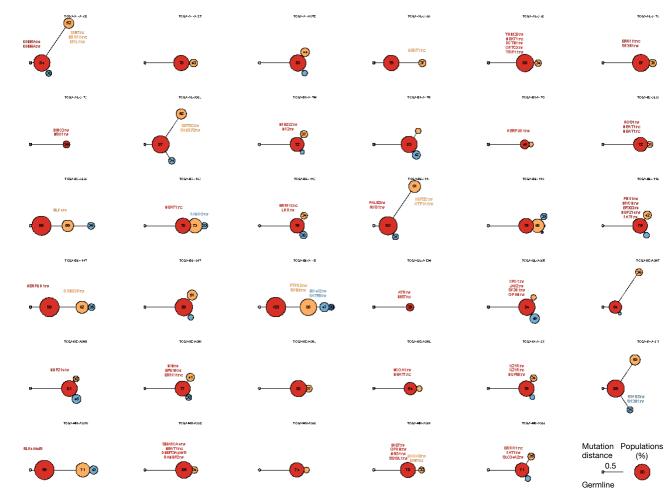
#### **Tumor Evolution: Highlight the Ordering of Key Mutations**



# **Construct evolutionary trees in pRCC**

- Infer mutation order and tree structure based on mutation abundance (PhyloWGS, Deshwar et al., 2015)
- Some of the key mutations occur in all the clones while others are just in some parts of the tree





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TOGA-ED-CERS

TCGA-06-116

TOGANIEASHE

TCGA45-5-67 9

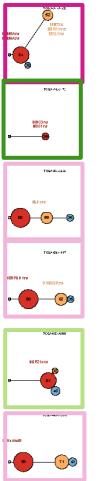
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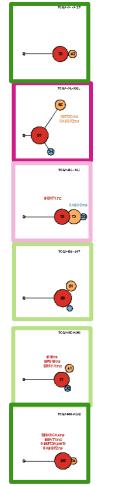
(%)

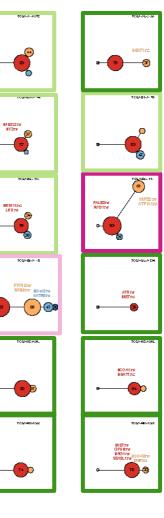
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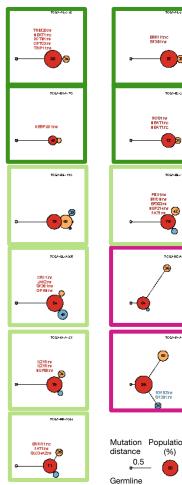
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ROSINS BEATING BEATING









TOGALAN 9 10F 82ns 8F381 75 Mutation Populations (%) 30

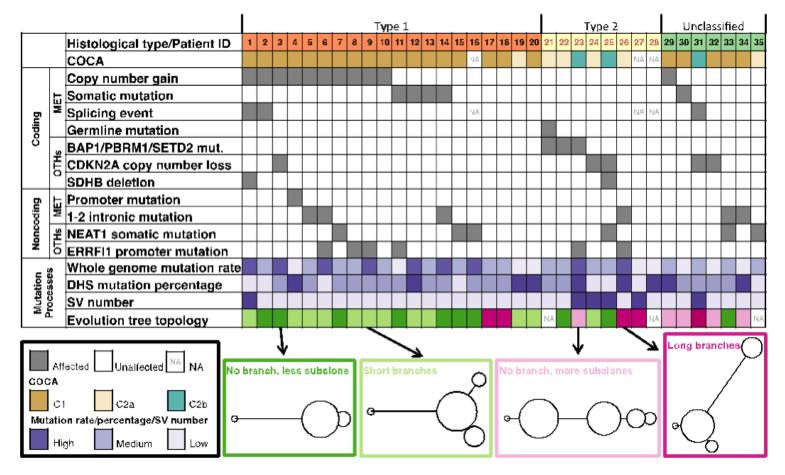
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TCGA-06- 110

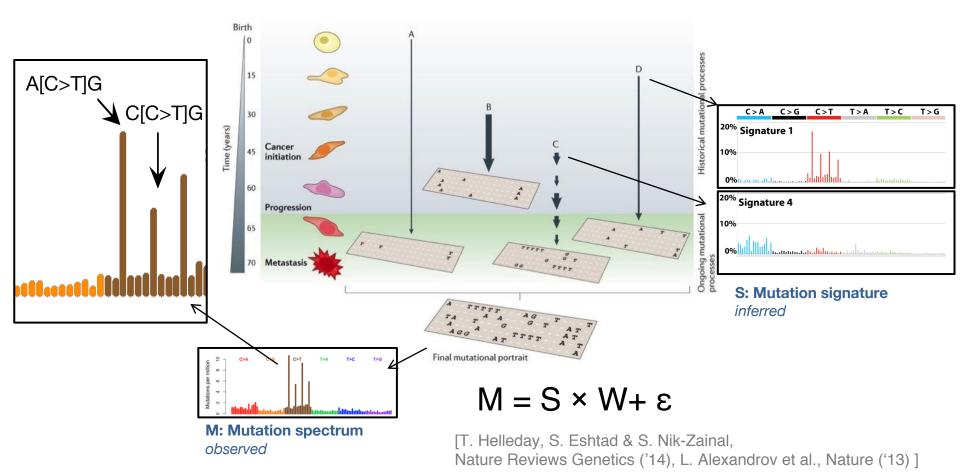
TOGALEASE

[S. Li, B. Shuch and M. Gerstein PLOS Genetics ('17)]

### Tree topology correlates with molecular subtypes

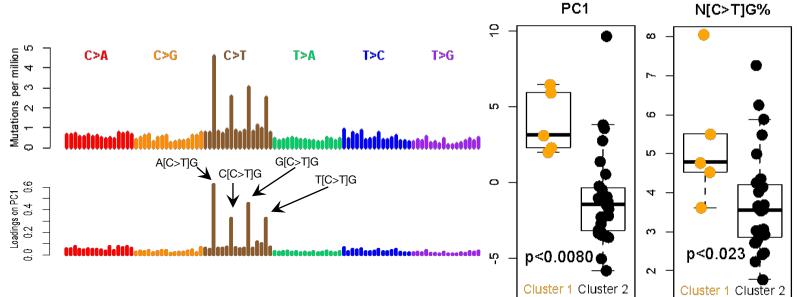


### Mutational processes carry context-specific signatures

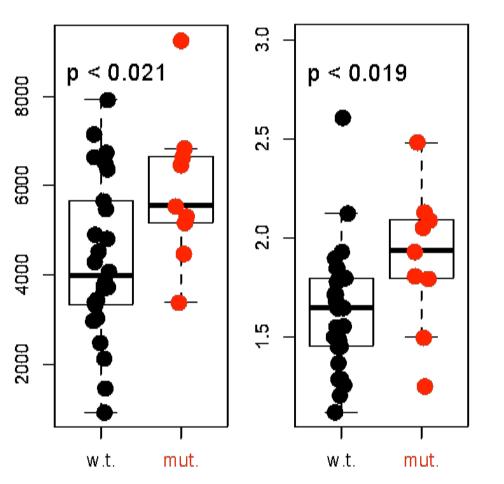


# CpGs drive inter-patient variation in pRCC mutational spectra

- The loadings on PC1 are mostly [C>T]G
- Confirmed by higher C>T% in CpGs in the hypermethylated group (cluster1)



DHS mutation %



Key mutation affects mutational landscape which, in turn, affects overall burden in pRCC

 Chromatin remodeling defect ("mut") leads to more mutations in open chromatin (raw number & fraction) in those pRCC cases w/ the mutation Prioritizing Variants in Personal Genomes: Using functional impact & recurrence, with particular application to cancer

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github.com/gersteinlab/**Frustration** S **Kumar**, D Clarke

github.com/gersteinlab/**MrTADfinder** KK **Yan**, S Lou

VAT.gersteinlab.org

L **Habegger**, S Balasubramanian, DZ Chen, E Khurana, A Sboner, A Harmanci, J Rozowsky, D Clarke, M Snyder

#### ALoFT.gersteinlab.org S Balasubramanian, Y Fu,

M Pawashe, P McGillivray, M Jin, J Liu, K Karczewski, D MacArthur

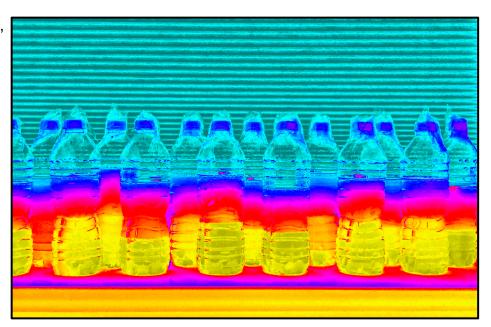
FunSeq.gersteinlab.org Y Fu, E Khurana, Z Liu, S Lou, J Bedford, XJ Mu, KY Yip

pRCC - S Li, B Shuch

**CostSeq2** - P **Muir**, S Li, S Lou, D Wang, DJ Spakowicz, L Salichos, J Zhang, GM Weinstock, F Isaacs, J Rozowsky

LARVA.gersteinlab.org L Lochovsky, J Zhang, Y Fu, E Khurana

MOAT.gersteinlab.org - L Lochovsky, J Zhang





### Info about this talk

### No Conflicts

Unless explicitly listed here. There are no conflicts of interest relevant to the material in this talk

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